

HUMAN OCULOMOTOR CONTROL SIMULATION

Carter Compton Collins, Ph.D.
Smith-Kettlewell Institute and Department of Visual Sciences,
University of the Pacific, San Francisco, California, 94115

ABSTRACT

Techniques have been developed for monitoring human oculomotor innervation and unrestrained muscle forces during large eye movements. Results of these measurements have led to a better understanding of eye movement control strategies and an Interactive Simulation Language (ISL) model of the nonlinear peripheral oculomotor plant which duplicates both healthy and pathological eye movement performance.

This model matches physiological, nonlinear innervational and viscous properties of the oculomotor plant for large excursions, performing satisfactorily over the entire + or -50° range of eye movement. The model duplicates the family of muscle length-tension curves, traces the static locus and properly generates agonist and antagonist muscle forces observed in freely moving eyes during tracking and saccadic movements. It duplicates dynamic saccadic length-tension loops (the most critical test of model performance). It shows the effect of muscle shortening on forces during a saccade, and the initial force increase in a relaxing antagonist.

The model simulates both physiological and pathological human eye movement performance. For example, the tropia, overshoot, velocity, and eye movement magnitudes are each duplicated for lateral rectus palsy patients. In addition, the effects of internuclear ophthalmoplegia are likewise duplicated. This striking identity of model performance with known pathology has important bearing on the possible clinical uses of models which can be evaluated from patient measurements.

An individual patient clinical model may eventually be able to qualitatively and quantitatively aid the ophthalmologist in his diagnosis and choice of surgical treatment plan. Operation on the model would permit the surgeon to compare various amounts and types of strabismus surgery and choose an optimum approach from many model responses before going to the operating room.

INTRODUCTION

When a person is unable to direct both eyes at the same object his visual disorder is called strabismus. The clinical correction of strabismus generally involves surgically moving the point of attachment of one or both of the extraocular muscles to realign the eyes. However, some 20 to 40 percent of all strabismus surgical patients must return for reoperation(1). Thus, there is clearly a need for a better understanding of the mechanical defects of strabismus and the surgery applied to overcome them.

METHODS

Computer simulation has become necessary for achieving a more complete and quantitative under-

standing of the many complex interrelationships involved in oculomotor control. Each anatomical element of the oculomotor plant must be separately represented since pathology and surgery affect each separately. Pathological ocular deviations are often significantly large, persisting to a great, but variable degree, over the entire range of ocular motility. Thus, a clinically useful model must be able to correctly simulate large eye movements embracing the performance of the grossly nonlinear oculomotor system over the complete range of plus or minus 50 degrees. The major nonlinearity is found in the relationship between the magnitude of innervation (the input control signal) and the position of the eye(2). In addition, muscle viscosity varies as muscle tension and is a nonlinear function of eye movement velocity.

The techniques for measurement of these anatomical and physiological entities have been described elsewhere(3,4) and hence are only briefly outlined here. First, the muscle innervation, or input control signal, θ , has been measured utilizing a specially developed multiple microelectrode technique for statistically sampling the simultaneous activity of up to 50 widely spaced, single motor units in a human oculorotary muscle during natural eye movements(3). This is the equivalent of sampling the message on the oculomotor nerve as sent from the brain to the eye muscle. By this means we have found that the brain strategy utilizes a nonlinear (square-law) pattern of input innervation to control the position of eye fixation. Next, to measure in situ muscle tension forces, T , under the natural conditions existing during unrestrained eye movements it was necessary to develop a miniature, high-performance, implantable recording strain gauge which has been separately described along with the observed patterns of human eye muscle forces(4). The elasticity, K , and viscosity, B , of the oculorotary muscles and passive globe (eyeball) restraining tissues have also been measured(5). Eye position, θ , is measured conventionally, either photoelectrically(6) or by means of electro-oculography(7).

A model based on these new human physiological findings has been evolved over the past few years and an analog computer simulation(2) has now been adapted to Interactive Simulation Language (ISL) for more powerful, extensive and flexible basic and clinical applications. This model of the oculomotor plant and the differential equations describing eye movements are shown in Fig. 1. The model reflects our physiological findings that each eye muscle pair functions anatomically in a reciprocally innervated, class A, "push-pull" mode which compensates for (i.e., linearizes) the square law input innervation. This results in a linear muscle force difference acting on the

predominantly linear globe restraining elasticity. The end result is that the eye movement system appears to be linear when observed from the outside. However, you are aware of the more than linear effort required to hold your eyes at the extreme gaze position of 50° left or right.

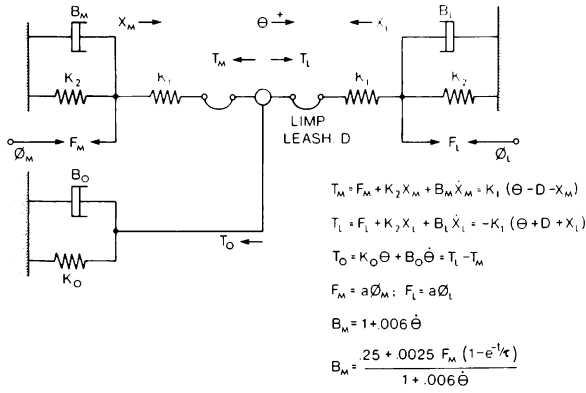


Fig. 1 - Diagram of the mechanical model of the human oculomotor plant and its differential equations of motion.

The physiologically measured input controls for this oculomotor plant model are represented in Fig. 2 as the static levels of innervation to the agonist and antagonist eye muscles, ϕ_M and ϕ_L . This transfer function relating the voluntary or desired eye position, ψ , to the level of static innervation, ϕ , is the result of the strategy chosen by the brain to control the steady-state or fixation position of the eyes.

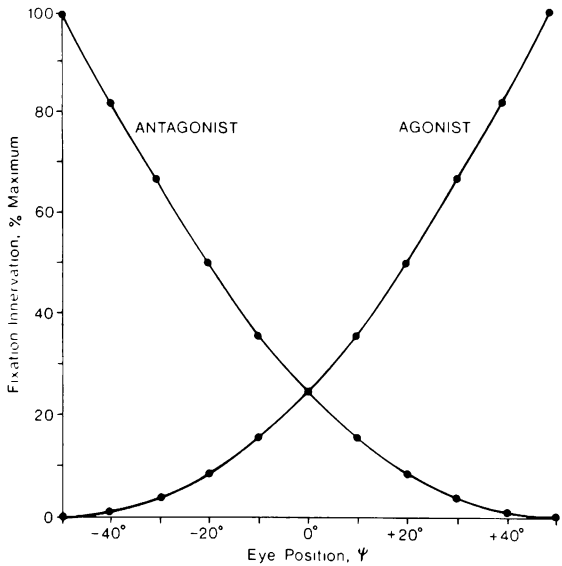


Fig. 2 - Steady-state input control signals (agonist and antagonist eye muscle innervation) as a function of eye position.

Superimposed upon this steady-state innervation are the transient control signals responsible for dynamic eye movements. The fastest eye movements are called saccades. These are quick, stepwise, coordinated movements of the eyes which are controlled by intricate, pre-programmed calculations of the central nervous system and result in remarkably accurate and rapid, precisely controlled flicking of the direction of gaze from one point to another. Figure 3 shows the magnitude and shape of the transient oculomotor control signals which produce saccades. These data have been hand averaged to remove the usual biological noise. Note a characteristic 10 msec. rise time to peak activity which is maintained for approximately one-half of the saccadic duration and falls with a time constant having a value of approximately one-fifth of the saccadic duration. The exact nature of the instantaneous value of this eye movement control signal is important in determining the transient time course of eye movements during saccades where large, inconstant and nonlinear viscous forces predominantly dictate movement.

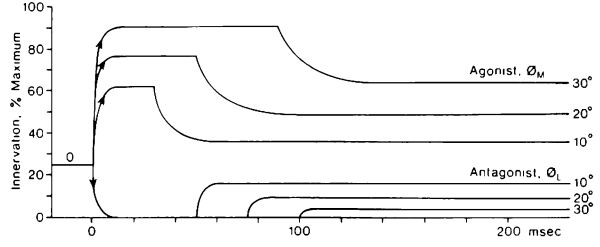


Fig. 3 - The time-varying pattern of the dynamic control signals of the eye muscles. These signals are superimposed on the steady-state levels to produce fast eye movements called saccades.

These physiologically measured, complex human eye movement control signals are used as inputs to drive the ISL model of the human oculomotor plant shown in the block diagram, Fig. 4. The left hand portion of the diagram performs the function of the central nervous system in generating these complex input control signals. In the upper left, ψ_0 represents the initial desired voluntary eye position (for example, straight ahead, the so-called primary position). ψ_1 then represents the desired new position of the eyes in degrees. The summer and multiplier in the upper and lower left portions of the diagram generate the steady-state innervation functions of Fig. 2. The left central elements of this model generate the transient components of the saccadic eye movement control signal which are added to the static components resulting in the complete eye movement control signals ϕ_M and ϕ_L .

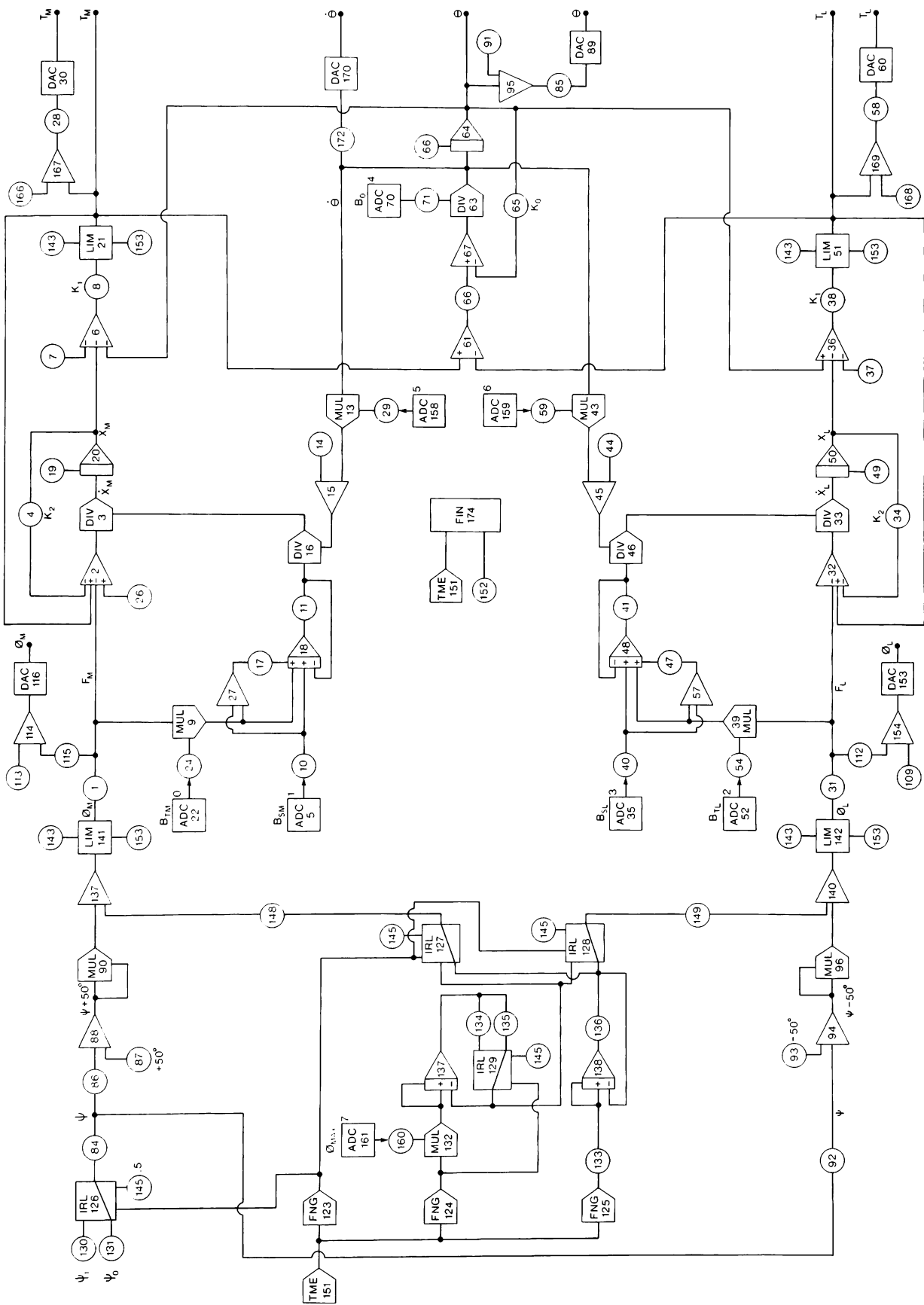


FIG. 4 - BLOCK DIAGRAM OF THE OCULOMOTOR PLANT

Fig. 5 - ISL listing of the oculomotor plant simulation programmed for 0 to 20° right saccade of left eye.

HUMAN OCULOMOTOR CONTROL SIMULATION

0	CON	0	= 5.000E-3	0
1	POT	141, 1.500E-2	= 3.749E 1	1
2	ADD	-1, -4, 21, 26	= 1.107E 0	2
3	DIV	2, 16	= 2.844E 0	3
4	POT	20, 1.200E 0	= -2.931E 1	4
5	ADC	1	= 1.113E 0	5
6	ADD	-7, -20, -64	= 3.713E 0	6
7	CON	7	= 2.000E 1	7
8	POT	6, 2.500E 0	= 9.281E 0	8
9	MUL	1, 24	= 1.062E-1	9
10	POT	5, 2.500E-1	= 2.783E-1	10
11	POT	18, 3.000E 1	= 3.838E-1	11
12	POT	3, 0.000E 0	= 0.000E 0	12
13	MUL	63, 29	= -1.397E-2	13
14	CON	14	= 1.000E 0	14
15	ADD	13, 14	= 9.860E-1	15
16	DIV	163, 15	= 3.892E-1	16
17	POT	27, 1.000E-2	= 3.840E-3	17
18	INT	17, 9, 10, -11	= 1.280E-2	18
19	CON	19	= -2.500E 1	19
20	INT	19, 3	= -2.442E 1	20
21	POT	8, 1.000E 0	= 9.281E 0	21
22	ADC	0	= 1.133E-1	22
23	CON	23	= 3.000E 2	23
24	POT	22, 2.500E-2	= 2.832E-3	24
25	POT	16, 1.000E 3	= 3.892E 2	25
26	POT	0, 0.000E 0	= 0.000E 0	26
27	ADD	9, 10	= 3.845E-1	27
28	POT	167, 9.000E-1	= 2.635E 1	28
29	POT	158, 2.500E-3	= 5.224E-3	29
30	DAC	28, 118, 64	= 0.328E-1	30
31	POT	142, 1.500E-2	= 3.750E 1	31
32	ADD	-31, -34, 51	= -1.071E 0	32
33	DIV	32, 46	= -2.804E 0	33
34	POT	50, 1.200E 0	= -2.745E 1	34
35	ADC	3	= 1.088E 0	35
36	ADD	-37, -50, 64	= 3.593E 0	36
37	CON	37	= 2.000E 1	37
38	POT	36, 2.500E 0	= 8.982E 0	38
39	MUL	31, 54	= 1.044E-1	39
40	POT	35, 2.500E-1	= 2.720E-1	40
41	POT	48, 3.000E 1	= 3.765E-1	41
42	POT	33, 0.000E 0	= 0.000E 0	42
43	MUL	63, 59	= -1.410E-2	43
44	CON	44	= 1.000E 0	44
45	ADD	43, 44	= 9.859E-1	45
46	DIV	165, 45	= 3.818E-1	46
47	POT	57, 1.000E-2	= 3.763E-3	47
48	INT	47, 39, 40, -41	= 1.255E-2	48
49	CON	49	= -2.500E 1	49
50	INT	49, 33	= -2.288E 1	50
51	POT	38, 1.000E 0	= 8.982E 0	51
52	ADC	2	= 1.113E-1	52
53	CON	53	= 3.000E 2	53
54	POT	52, 2.500E-2	= 2.783E-3	54
55	POT	46, 1.000E 3	= 3.818E 2	55
56	CON	56	= 0.000E 0	56
57	ADD	39, 40	= 3.763E-1	57
58	POT	169, 9.000E-1	= 2.608E 1	58
59	POT	159, 2.500E-3	= 5.259E-3	59
60	DAC	58, 118, 64	= 0.326E-1	60
61	ADD	21, -51	= 2.993E-1	61
62	CON	62	= 0.000E 0	62
63	DIV	67, 71	= -2.662E 0	63
64	INT	62, 63	= 7.097E-1	64
65	POT	64, 5.000E-1	= 3.549E-1	65
66	POT	61, 1.000E 0	= 2.993E-1	66
67	ADD	66, -65	= -5.554E-2	67
68	POT	67, 5.000E 0	= -2.777E-1	68
69	POT	64, 5.000E 0	= 3.549E 0	69
70	ADC	4	= 8.379E-1	70
71	POT	70, 2.500E-2	= 2.095E-2	71
72	CON	72	= 0.000E 0	72
73	CON	73	= 0.000E 0	73
74	CON	74	= 0.000E 0	74
75	CON	75	= 0.000E 0	75
76	CON	76	= 0.000E 0	76
77	CON	77	= 0.000E 0	77
78	CON	78	= 0.000E 0	78
79	CON	79	= 0.000E 0	79
80	POT	122, 1.000E 0	= 0.000E 0	80
81	POT	80, 1.000E 0	= 0.000E 0	81
82	CON	82	= 0.000E 0	82
83	INT	82, 81, -84	= -3.161E 2	83
84	POT	126, 1.000E 0	= 0.000E 0	84
85	POT	95, 8.400E-1	= 9.000E 0	85
86	POT	84, 1.000E 0	= 0.000E 0	86
87	CON	87	= 5.000E 1	87
88	ADD	86, 87	= 5.000E 1	88
89	DAC	85, 119, 64	= 0.112E-1	89
90	MUL	88, 88	= 2.500E 3	90
91	CON	91	= 1.000E 1	91
92	POT	84, 1.000E 0	= 0.000E 0	92
93	CON	93	= 5.000E 1	93
94	ADD	92, -93	= -5.000E 1	94
95	ADD	64, 91	= 1.071E 1	95
96	MUL	94, 94	= 2.500E 3	96
97	CON	97	= -0.000E 0	97
98	CON	98	= 1.000E-2	98
99	CON	99	= 0.000E 0	99
100	POT	103, 1.500E-2	= 1.960E 59	100
101	CON	101	= -5.000E 1	101
102	ADD	100, 101	= 1.960E 59	102
103	TDL	106, 104, 105	= 1.307E 61	103
104	CON	104	= 1.000E 0	104
105	CON	105	= 2.500E 2	105
106	POT	151, 1.000E 0	= 1.500E 0	106
107	TDL	110, 108, 109	= 2.774E 63	107
108	CON	108	= 0.000E 0	108
109	CON	109	= 5.000E 1	109
110	CON	110	= 0.000E 0	110
111	CON	111	= 0.000E 0	111
112	POT	31, 1.000E-1	= 3.750E 0	112
113	CON	113	= 5.000E 1	113
114	ADD	113, 115	= 5.375E 1	114
115	POT	1, 1.000E-1	= 3.749E 0	115
116	DAC	114, 118, 64	= 0.671E-1	116
117	CON	117	= 1.000E 0	117
118	CON	118	= 1.000E 2	118
119	CON	119	= 1.000E 2	119
120	CON	120	= 0.000E 0	120
121	CON	121	= 0.000E 0	121
122	CON	122	= 0.000E 0	122
123	FNG	151	= 0.000E 0	123
		0.000E 0	0.000E 0	
		4.900E-2	0.000E 0	
		5.000E-2	1.000E 0	
		5.000E-1	1.000E 0	
		5.010E-1	0.000E 0	
		1.000E 0	0.000E 0	
		1.000E 1	0.000E 0	
124	FNG	151	= 0.000E 0	124
		0.000E 0	0.000E 0	
		4.900E-2	0.000E 0	
		5.000E-2	1.000E 0	
		1.000E-1	1.000E 0	
		1.010E-1	0.000E 0	
		1.000E 0	0.000E 0	
		1.000E 1	0.000E 0	
125	FNG	151	= 0.000E 0	125
		0.000E 0	0.000E 0	
		4.900E-2	0.000E 0	
		5.000E-2	-1.000E 0	
		1.250E-1	-1.000E 0	
		1.260E-1	0.000E 0	
		1.000E 0	0.000E 0	
		1.000E 1	0.000E 0	
126	IRL	123, 145, 130, 131	= 0.000E 0	
127	IRL	123, 145, 137, 136	= -3.773E-204	
128	IRL	123, 145, 136, 129	= 7.634E-32	
129	IRL	124, 145, 134, 135	= 6.773E-32	
130	CON	130	= 2.000E 1	130
131	CON	131	= 0.000E 0	131
132	MUL	124, 160	= 0.000E 0	132
133	POT	125, 1.200E 3	= 0.000E 0	133
134	POT	137, 2.500E 2	= 3.005E-31	134
135	POT	137, 5.000E 1	= 6.011E-32	135
136	POT	138, 2.500E 2	= -1.886E-204	136
137	INT	132, 132, -129	= 1.067E-33	137
138	INT	133, 133, -136	= -3.773E-207	138

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139 ADD 90, 148 = 2.500E 3 139
140 ADD 96, 149 = 2.500E 3 140
141 LIM 143, 153, 139 = 2.500E 3 141
142 LIM 143, 153, 140 = 2.500E 3 142
143 CON 143 = 1.000E 5 143
144 DAC 154, 118, 64 = 0.671E-1 144
145 CON 145 = 5.000E-1 145
146 IRL 123, 145, 132, 147 = 0.000E 0 146
147 POT 124, 5.000E 1 = 0.000E 0 147
148 POT 127, 2.000E 0 = -7.546E-204 148
149 POT 128, 1.000E 0 = 7.634E-32 149
150 CON 150 = 0.000E 0 150
151 TME 151 = 1.502E 0 151
152 CON 152 = 1.500E 0 152
153 CON 153 = 0.000E 0 153
154 ADD 109, 112 = 5.375E 1 154
155 CON 155 = 0.000E 0 155
156 CON 156 = 0.000E 0 156
157 CON 157 = 1.000E 3 157
158 ADC 5 = 2.090E 0 158
159 ADC 6 = 2.105E 0 159
160 POT 161, 1.875E 5 = 5.830E 5 160
161 ADC 7 = 3.123E 0 161
162 INT 11, 11, -163 = 1.279E-2 162
163 POT 162, 3.000E 1 = 3.838E-1 163
164 INT 41, 41, -165 = 1.255E-2 164
165 POT 164, 3.000E 1 = 3.765E-1 165
166 CON 166 = 2.000E 1 166
167 ADD 21, 166 = 2.928E 1 167
168 CON 168 = 2.000E 1 168
169 ADD 51, 168 = 2.898E 1 169
170 DAC 172, 118, 64 = 0.000E 0 170
171 DAC 151, 117, 128 = 1.249E-1 171
172 POT 63, 2.500E-2 = -6.654E-2 172
173 DAC 143, 143, 64 = 1.249E-1 173
174 FIN 151, 152 = 0.000E 0 174
175 END 0 = -7.831E 1227 175

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The upper and lower right elements of the model represent the medial and lateral rectus respectively. The contractile element consists of two dividers, a multiplier and an integrator to represent respectively the nonlinear muscle viscosity as a function of developed tension, eye movement velocity and the temporal dynamics of the contractile element. The series elastic element, K_1 of each muscle is on the right of the collection of muscle elements. The muscle outputs consist of tendon tensions, T_M and T_L .

These muscle tensions are algebraically added and applied to the globe dynamics, K_0 and B_0 , in the right central portion of the model. The resulting output is seen as eye position, θ , and eye movement velocity, $\dot{\theta}$.

Figure 5 is an ISL listing of the oculomotor plant model described here.

RESULTS

Figure 6 shows the model output performance characteristics on the length-tension plane as reproduced directly from a storage oscilloscope. Figure 7 is a hand-traced rendition of these curves labeled for greater clarity. These data agree quite well with recorded physiological measurements on human oculorotary muscle patients. In fact, Fig. 7 can serve as a summary of the measured physiological output of the normal human oculomotor plant⁽²⁾ showing both the steady-state and dynamic performance of this system on the length-tension plane. Muscle length increases to the right and is shown in terms of eye position.

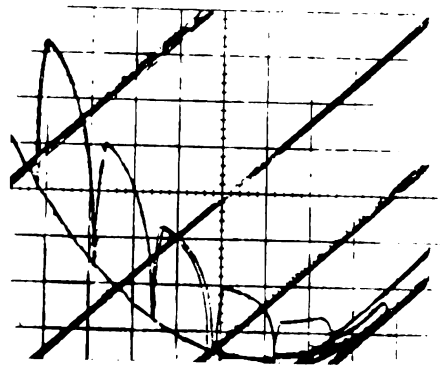


Fig. 6 - Static and dynamic performances of the model on the length-tension plane taken directly from a storage oscilloscope output.

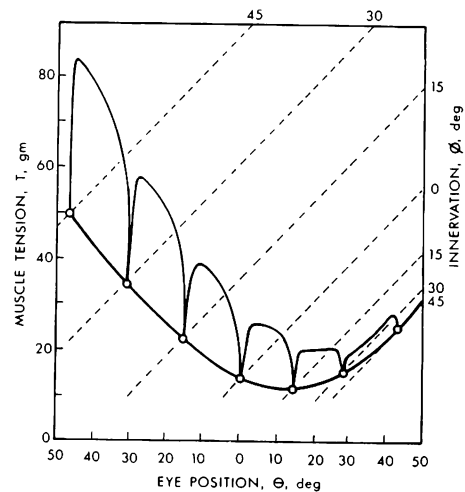


Fig. 7 - Labeled drawing of the model outputs of Fig. 6 showing length-tension characteristics of a muscle, the static locus and dynamic saccadic loops.

Right gaze is to the right on the diagram. The dashed lines in this figure represent the family of length-tension curves of the left lateral rectus muscle as a function of various degrees of fixed innervation shown as the parameter on the right hand ordinate. Zero indicates an innervation for zero degrees, or straight ahead; progressing upward the numbers represent increasing innervation to the labeled positions of gaze into the muscle's field of action, in this case, left gaze. Downward, innervation is decreasing to the labeled positions of right gaze out of the muscle's field of action. Note that these curves are not evenly spaced reflecting the nonlinearity of input innervation.

The heavy curved line intersecting the small circles at the bottom of the diagram indicates the measured tensions utilized by the oculomotor system to hold the eye at any degree of eccentric gaze. We call this curve the static locus of fixation tensions. Tensions recorded during smooth following movements of the eyes fall on or slightly above this static locus.

Typical dynamic length-tension loops for 15° fast saccadic movements of the eyes produced by the agonist (active) muscle are shown in Figs. 6 and 7 as counterclockwise-traced force pulses extending between the open circles on the static locus. These circles represent the starting (right) and finishing (left) points of saccadic eye position. [A typical dynamic length-tension loop for the antagonist (relaxing) muscle would be shown as a much smaller, flattened clockwise path moving to the right between the initial (left) and final (right) positions of the eye.] The tops of the saccadic force peaks in Fig. 7 define an upper curved line which represents the maximum forces recorded during any eye movement. The area between this upper curve and the static locus we call the operational envelope. This operational envelope defines the normal ranges of tensions for horizontal eye movements as measured during the waking state. The operational envelope of normal eye movement forces measured at the tendon of the muscle is determined by neuromuscular control, the innervational strategy of the central nervous system operating on the mechanical viscoelastic characteristics of the muscles and globe. The combination of these factors restricts normal muscle activity to a mere 20 percent of the area of the length-tension diagram. All areas on the length-tension diagram outside the operational envelope are forbidden regions during normal eye movements. Only during abnormal or pathological conditions do the muscle forces leave the operational envelope area of the length-tension diagram. Thus, the operational envelope has clinical significance relating to diagnosis utilizing forced duction or other techniques.

In the model of Figs. 1 and 4, the contractile element force consists of three components: the developed force, F , due to innervation, ϕ ; the force-velocity relationship of the muscle, or viscosity, B ; and the force due to the parallel elastic element, K_2 , contributing to the length-tension characteristic of muscle. The sum of these three forces is equal to the force measured at the muscle tendon. However, any one component of this contractile element force cannot be uniquely determined at the tendon because of the isolating series elastic element, K_1 .

Figure 8 is a time based record of the performance of the model left eye in response to a zero to 20-degree right saccadic input innervation, ϕ_M of the medial rectus muscle shown on the top channel of the oscillographic record. The next lower record shows the innervation, ϕ_L of the antagonist lateral rectus muscle which was reciprocally innervated, in this case turned completely off during the saccade in accord with the quantitative results of physiological measurements of innervation.

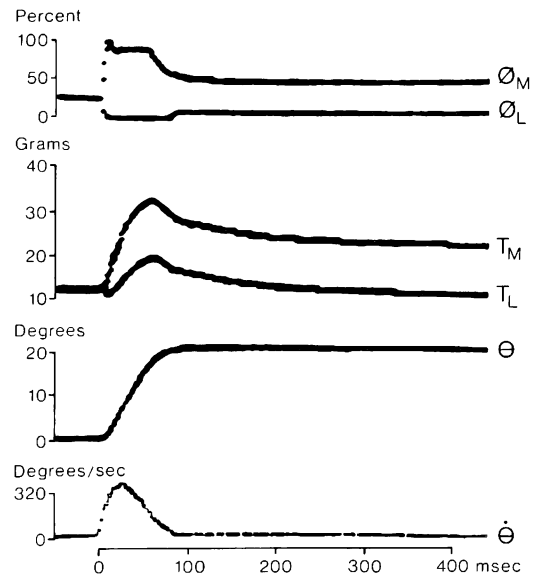


Fig. 8 - Dynamic performance of the model as a function of time in response to the reciprocal innervation, ϕ_M and ϕ_L (top traces) for a zero to 20° right saccade of the left eye. The middle traces are muscle tensions, T_M and T_L , and the lower traces are eye position, θ , and velocity, $\dot{\theta}$.

The next two records are those of model-derived muscle tensions, T_M and T_L , of the medial and lateral rectus muscles respectively. The agonist medial rectus muscle force is seen to be greater, peaking in about 50 msec and then slowly decaying to its steady-state fixation or holding value. Just below it, the lateral rectus muscle tension is also seen to increase before decreasing to its final steady-state value. This increase was unexpected since the lateral rectus muscle innervation drops to zero during the saccade, and separate isometric measurements of this antagonist muscle force have shown that the muscle force drops below baseline tensions. The reason for the muscle force increase is that this antagonist muscle is stretched by the agonist muscle at a rate which causes the antagonist muscle viscous forces to increase more rapidly and with greater magnitude than the force decrease due to the drop in innervation. Both the agonist and antagonist model muscle forces faithfully reproduce the patterns and magnitude of forces as measured by our "C" gauges implanted in human oculomotor muscles during strabismus surgery(4).

The next lower record represents the resultant eye movement, θ , which again truly represents the pattern of eye movement seen in normal human subjects. Finally, the lowest record shows the eye movement velocity, $\dot{\theta}$, which again is a sensitive test of model performance. This velocity profile agrees quite well with measured human eye movement data and again illustrates the realistic performance of this oculomotor plant simulation.

- Fig. 9A - Oculomotor model simulation of a normal left eye.
- 9B - Simulation of a pathological eye movement with a paralyzed antagonist left lateral rectus muscle.
- 9C - Simulated pathological eye movement with a paralyzed agonist left medial rectus muscle.
- 9D - Actual patient records of a lateral rectus palsy patient. Note how closely these are duplicated by the model.

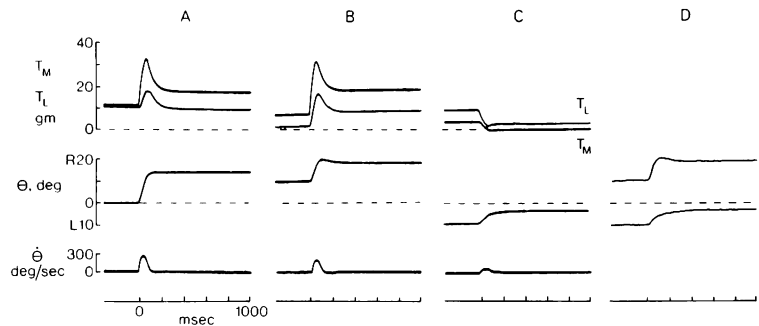


Figure 9 compares pathological oculomotor performance with the normal performance record of Fig. 8. Figure 9A is an oculomotor simulation of a normal left eye during a 0 to 15-degree right saccade. At the top are the unrestricted medial and lateral muscle tensions, T_M and T_L , shown on the same scale to illustrate comparative performance. The next lower trace shows eye position, θ . The bottom trace indicates velocity, $\dot{\theta}$.

Figure 9B shows the model simulation of a pathological left eye during a 15° right saccade with a paralyzed antagonist left lateral rectus muscle. Note the 10° right esotropia (inward misalignment), 3° saccadic overshoot, 150° per second velocity (about half normal), and 8° final eye movement instead of the normal 15°.

Figure 9C shows the model simulation of a pathological left eye movement during a 15° right saccade with a paralyzed agonist left medial rectus muscle. Note the 10° left exotropia (outward misalignment), 500 msec saccadic duration, 60° per second velocity, and 7° final eye movement.

Figure 9D presents eye movement records of a lateral rectus palsy patient. The top record with a palsied antagonist shows the tropia, overshoot, velocity, and size of eye movement which are duplicated by the model in Fig. 9B. The bottom trace is a patient record of movement with a palsied muscle acting as agonist. Note the model duplicates this pathological eye movement in Fig. 9C.

DISCUSSION

It can be seen that the model simulates both physiological and pathological human eye movement performance. This striking identity of model performance with known pathology has important bearing on the possible clinical uses of models which can be evaluated from patient measurements. By making simple clinical office measurements with a strain gauge forceps it appears to be possible to evaluate the various model elements of the patient's oculomotor plant in the surgeon's office. Calibrated forceps measurements⁽⁸⁾ of isometric forces during contralateral eye fixations can determine neuromuscular functional imbalances; and contractures or mechanical restrictions may be quantitatively determined in the office with position monitored forceps measurements of ocular length-tension characteristics.

An individual patient clinical model thus resulting may be able to quantitatively and qualitatively aid the ophthalmologist in his diagnosis and choice of surgical treatment plan. An interactive graphic CRT display now under development in our laboratories would provide immediate feedback indicating the results of various combinations of simple surgical techniques on oculomotor performance and alignment. For example, the effects of surgery on comitance (relative alignment of the eyes) could be instantly and directly displayed in graphical form. Simulated surgery on the model would permit an ophthalmologist to explore various types and amounts of corrective surgery. He would then have the choice of pursuing a conventional plan or of considering a different surgical approach suggested by newly derived computer data based on quantitative measurements of the patient before going to surgery.

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